1. **COURSE ID:** BIOL 230  
**TITLE:** Introductory Cell Biology  
**C-ID:** BIOL 190  
**Units:** 4.0 units  
**Hours/Semester:** 48.0-54.0 Lecture hours; and 48.0-54.0 Lab hours  
**Method of Grading:** Letter Grade Only  
**Prerequisite:** CHEM 210, and either BIOL 210 or BIOL 220 or BIOL 215 (Skyline) or BIOL 225 (Canada)  
**Recommended Preparation:** Eligibility for ENGL 100 or 105

2. **COURSE DESIGNATION:**  
**Degree Credit**  
**Transfer credit:** CSU; UC  
**AA/AS Degree Requirements:**  
CSM - GENERAL EDUCATION REQUIREMENTS: E5a. Natural Science  
CSU GE:  
CSU GE Area B: SCIENTIFIC INQUIRY AND QUANTITATIVE REASONING: B2 - Life Science  
CSU GE Area B: SCIENTIFIC INQUIRY AND QUANTITATIVE REASONING: B3 - Laboratory Activity  
IGETC:  
IGETC Area 5: PHYSICAL AND BIOLOGICAL SCIENCES: B: Biological Science  
IGETC Area 5: PHYSICAL AND BIOLOGICAL SCIENCES: C: Science Laboratory

3. **COURSE DESCRIPTIONS:**  
**Catalog Description:**  
Evaluation and analysis of the living cell and its components. Examines cell structures and metabolism as they relate to cell function and reproduction. (Recommended for all life science and medical science majors.) One or more field trips may be required. Extra supplies may be required.

4. **STUDENT LEARNING OUTCOME(S) (SLO’S):**  
Upon successful completion of this course, a student will meet the following outcomes:  
1. Describe and relate the origin of life and subsequent evolution of organelle structure and function in prokaryotic and eukaryotic cells, including landmark experiments.  
2. Identify and describe structure and major functions of cellular organic molecules.  
4. Distinguish and compare biochemistry, synthesis of nucleic acids and proteins, including major experimental techniques.  
5. Explain and apply principles of classical/Mendelian Genetics to problems in genetics or biotechnology.  
7. Demonstrate basic laboratory skills for objective investigation of cell biology phenomena and a scientific approach to investigating cells.  

5. **SPECIFIC INSTRUCTIONAL OBJECTIVES:**  
Upon successful completion of this course, a student will be able to:  
1. Describe and relate the origin of life and subsequent evolution of organelle structure and function in prokaryotic and eukaryotic cells, including landmark experiments.  
2. Identify and describe structure and major functions of cellular organic molecules.  
4. Distinguish and compare biochemistry, synthesis of nucleic acids and proteins, including major experimental techniques.  
5. Explain and apply principles of classical/Mendelian Genetics to problems in genetics or biotechnology.  
7. Demonstrate basic laboratory skills for objective investigation of cell biology phenomena and a scientific approach to investigating cells.  
6. COURSE CONTENT:

Lecture Content:

Course Content-lecture topics
1. Origin of Life; evolution of organelle structure and function in prokaryotic and eukaryotic cells; Eukaryotic Cell Organelles: structure, functions (SLO #1)
2. Cell division, cell cycle control; Cell fractionation and analysis (SLO #1)
3. Water and organic molecules in cells; complex structure and function of proteins (SLO #2)
4. Principles of Metabolism: endergonic and exergonic reactions, activation energy, energy coupling, enzyme catalysis (SLO #3)
5. Membranes: phospholipid bilayer, role of proteins in transport, communication, energy requirements for transport (SLO #1, 2, 3)
6. Cellular Respiration: overall energy considerations, oxidation-reduction cycles, locations, glycolysis, fermentation, citric acid cycle, electron transport, role of mitochondrial membrane, chemiosmotic mechanism of ATP synthesis; allosteric regulation of enzymes in metabolism (SLO #3)
7. Photosynthesis: overall energy considerations, oxidation-reduction cycles, role of chlorophyll, photosystems, thylakoid membranes, photophosphorylation mechanism (SLO #3)
8. DNA Structure, Replication: evidence for semiconservative mechanism; process at chromosomal and biochemical levels (SLO #4)
9. RNA structure and synthesis; Molecular (nucleic acid) hybridization (SLO #4)
10. Protein synthesis: genetic code, components, steps; Mutation causes and consequences and relation to classical/Mendelian genetics (SLO #4)
11. DNA Technology: restriction enzymes, cloning, analysis of DNA by hybridization, applications (SLO #4)
12. Prokaryotic Gene Regulation: coupled transcription, translation; operon regulation (SLO #6)
13. Eukaryotic Gene Structure, Regulation: introns, exons, RNA processing, steroid hormone action, chromosome packaging; comparison of prokaryotic and eukaryotic regulation; antibody genes (SLO #6)
14. Cell Communication: hormones, target cells, G-protein-linked receptors, cyclic AMP, and allosteric activation of protein kinase (SLO #6)

Lab Content:

Course content-laboratory topics
1. Pipets, dilutions, metric units (SLO #7, 8)
2. Molecular models (SLO #7, 8)
3. Protein extraction, quantification and analysis by electrophoresis (SLO #2, 7, 8)
4. Science and Cells: Designing Experiments to Test Hypotheses; review use of microscope (SLO #7, 8)
5. Bacterial Growth, selective plating (SLO #7, 8)
6. Enzyme Activity (SLO #2, 7, 8)
7. Modeling and molecular explanations of classical/Mendelian genetics (SLO #2, 5)
8. DNA extraction and analysis: mini-plasmid preparation, restriction enzyme digests, sizing DNA by electrophoresis (SLO #2, 7, 8)
9. Modeling principles of complementary base pairing and information flow: replication, transcription, DNA sequencing; Molecular basis for Mendelian inheritance (SLO #4, 7, 8)
10. Student models of cell biology processes or structures; presentations on cell biology topics (SLO #7, 8)

7. REPRESENTATIVE METHODS OF INSTRUCTION:

Typical methods of instruction may include:
A. Lecture
B. Lab
C. Activity
D. Directed Study
E. Discussion
F. Experiments
G. Guest Speakers
H. Observation and Demonstration

8. REPRESENTATIVE ASSIGNMENTS

Representative assignments in this course may include, but are not limited to the following:
Writing Assignments:
Written laboratory assignments clearly record each exercise: theoretical questions, experimental design, procedures, observations, summary and interpretation of results. Assignments evaluate how well results compare to expectations, relate experiments and results to principles studied in lecture.

Independent research into selected topics, resulting in poster, model and oral presentations demonstrating identification and evaluation of main concepts of topic.

Reading Assignments:
Reading in textbok to restate lecture and laboratory concepts, illustrate points of lecture, define terms, and provide examples.

Reading journal and website articles assigned to further research lab and lecture concepts.

Other Outside Assignments:
- Preparing 4 individual presentations of answers to assigned Study Questions (counted as part of lab attendance);
- Preparing 2 individual presentations of amino acid interactions in proteins with 3 x 5 card amino acid sets made in Lab 2 (counted as part of lab assignment grade);
- Designing and constructing a model of basic lecture concept with one other student (counted as part of lab assignment grade).

9. REPRESENTATIVE METHODS OF EVALUATION
Representative methods of evaluation may include:
A. Class Participation
B. Class Work
C. Exams/Tests
D. Homework
E. Lab Activities
F. Oral Presentation
G. Projects
H. Written examination
I. Midterm and Final exams assess student's ability to communicate SLOs 1-6, using SLO 7. Laboratory assignments assess student's methods, careful recording of observations, results, clear evaluation of conclusions, SLOs 6 and 7 and reinforcement of SLOs 1-5. Oral and poster or model presentations reveal students' ability to identify and evaluate main concepts of selected topics.

10. REPRESENTATIVE TEXT(S):
Possible textbooks include:
B. Hardin et al. BECKER'S WORLD OF THE CELL, 8th ed. Benjamin Cummings, 2012
Possible manuals include:
A. Diamond. Laboratory exercises are individually prepared and uploaded to WebAccess site for the course, WebAccess site at College of San Mateo, 08-01-2013

Origination Date: October 2014
Curriculum Committee Approval Date: January 2015
Effective Term: Fall 2015
Course Originator: Kathleen Diamond